## Communications to the Editor

## ISOLATION AND STRUCTURAL ELUCIDATION OF ANTIOXIDATIVE AGENTS, ANTIOSTATINS $A_1$ TO $A_4$ AND $B_2$ TO $B_5$

Sir:

Active oxygen species cause a variety of diseases such as ischemia-reperfusion, inflammation, autoimmune disease, diabetes, rheumatism, cardiovascular diseases and cancer-initiation<sup>1,2)</sup>. Thus, it could be expected that antioxidative agents may prevent these diseases.

During the course of a screening program for novel compounds showing antioxidant activity, we obtained a new naphthoquinone derivative naphterpin<sup>3)</sup>. Further screening resulted in the isolation of new antioxidative agents named antiostatins  $A_1$  to  $A_4$  and  $B_2$  to  $B_5$  from *Streptomyces cyaneus* 2007-SV<sub>1</sub>. They showed strong inhibitory activity against lipid peroxidation induced by free radicals in rat liver microsome preparations free from vitamin E<sup>4)</sup>. In this paper, we report the fermentation, isolation and structural studies of these new metabolites.

A stock culture of S. cyaneus 2007-SV<sub>1</sub> was inoculated into 500-ml Erlenmeyer flasks containing 100 ml of a seed medium (starch 2.5%, soybean meal 1.5%, dry yeast 0.2% and CaCO<sub>3</sub> 0.4%, pH 6.2 before sterilization) and incubated at 27°C for 2 days on a rotary shaker. The seed culture (600 ml) thus obtained was transferred into a 50-liter jar fermenter containing 30 liters of the same medium, and cultivation was carried out at 27°C for 48 hours under agitation at 400 rpm and aeration at 30 liters per minute.

The active materials were isolated according to the procedures shown in Scheme 1. Each component of the antiostatins was separated at the final stage by reversed phase HPLC (Senshu Pak C-18) and was obtained as a pale yellow powder. The antiostatins were soluble in MeOH, CHCl<sub>3</sub>, acetone and ethyl acetate, but insoluble in hexane and water. Based on the structural features of the substituent at C-4, they were divided into two groups, the A series and B series. Their structural studies were carried out using the main components  $A_1$  (yield 2.5 mg) and  $B_4$  (yield 2.0 mg) as explained in the following. Antiostatin A<sub>1</sub>: MP 180~183°C (dec); UV  $\lambda_{max}^{MeOH}$  nm ( $\epsilon$ ) 220 (28,500), 238 (26,200), 301 (16,100), 338 (4,250), 350 (4,250); IR  $v_{max}$  (KBr) cm<sup>-1</sup> 3350, 1620, 1585; HREI-MS (m/z) 324.1822 (M<sup>+</sup>), calcd for C<sub>20</sub>H<sub>24</sub>N<sub>2</sub>O<sub>2</sub>, 324.1838.

Antiostatin B<sub>4</sub>: MP 118~120 °C<sup>†</sup>; UV  $\lambda_{max}^{MOH}$  nm ( $\varepsilon$ ) 218 (44,300), 238 (35,300), 301 (20,900), 338 (6,160), 352 (6,290); IR  $\nu_{max}$  (KBr) cm<sup>-1</sup> 3480, 3370, 3340, 1690, 1670, 1550; HRFAB-MS (*m*/*z*) 452.2763 (M<sup>+</sup>), calcd for C<sub>26</sub>H<sub>36</sub>N<sub>4</sub>O<sub>3</sub>, 452.2787.

The UV spectral similarity between antiostatins  $A_1$  and  $B_4$ , and carbazomycin  $B^{5,6)}$  suggested the presence of the carbazole nucleus in these compounds. Their structures were determined by <sup>1</sup>H and <sup>13</sup>C NMR spectral analyses and by comparison

Scheme 1. Isolation and purification of antiostatins.

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Whole broth (60 liters, pH 7.5)
  filtered
Mycerial cake
  extracted with acetone
  concentrated in vacuo
  extracted with EtOAc
EtOAc layer
  evaporated to dryness
Silica gel column
  eluted with hexane - EtOAc (3:1)
  evaporated in vacuo
Sephadex LH-20
  eluted with CH<sub>3</sub>OH - CHCl<sub>3</sub> (1:1)
  evaporated to dryness
Toyopearl HW-40
  eluted with MeOH
  concentrated in vacuo
HPLC Senshu Pak C-18
   eluted with MeOH - H<sub>2</sub>O (81:19)
      A<sub>2</sub> A<sub>3</sub> A<sub>4</sub> B<sub>2</sub> B<sub>3</sub> B<sub>4</sub> B<sub>5</sub>
Α1
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<sup>†</sup> Antiostatins  $B_2$  to  $B_5$  gradually started to sublime at around 90°C.

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|--------------|----------------|----------------|------------------------|
| Carbon       | A <sub>1</sub> | B <sub>4</sub> | Carazostatin           |
| 1            | 123.3 (s)      | 123.7 (s)      | 124.1 (s) <sup>b</sup> |
| 2            | 125.8 (s)      | 126.0 (s)      | 121.4 (s)              |
| 3            | 144.3 (s)      | 143.9 (s)      | 148.2 (s)              |
| 4            | 118.3 (s)      | 117.6 (s)      | 103.0 (s)              |
| 4a           | 115.5 (s)      | 115.7 (s)      | 120.9 (s)              |
| 4b           | 123.8 (s)      | 123.7 (s)      | 123.7 (s)              |
| 5            | 122.8 (d)      | 122.6 (d)      | 120.0 (d)              |
| 6            | 119.2 (d)      | 119.7 (d)      | 118.9 (d)              |
| 7            | 125.8 (d)      | 126.0 (d)      | 125.2 (d)              |
| 8            | 111.8 (d)      | 112.1 (d)      | 110.6 (d)              |
| 8a           | 141.4 (s)      | 141.8 (s)      | 139.8 (s)              |
| 9a           | 135.1 (s)      | 135.3 (s)      | 134.0 (s)              |
| 10           | 14.6 (q)       | 13.3 (q)       | 12.0 (q)               |
| 1′           | 30.0 (t)       | 29.7 (t)       | 28.8 (t)               |
| 2'           | 30.1 (t)       | 31.1 (t)       | 29.5 (t)               |
| 3'           | 33.1 (t)       | 31.0 (t)       | 30.0 (t)               |
| 4′           | 23.5 (t)       | 30.6 (t)       | 29.3 (t)               |
| 5'           | 13.0 (q)       | 33.2 (t)       | 31.7 (t)               |
| 6'           |                | 23.8 (t)       | 22.7 (t)               |
| 7′           | _              | 14.9 (q)       | 14.1 (q)               |
| 1″           | 172.1 (s)      | 155.9 (s)      |                        |
| 2″           | 23.7 (q)       | 156.8 (s)      |                        |
| 3″           |                | 48.3 (t)       |                        |
| 4″           |                | 30.1 (d)       |                        |
| 5″           |                | 20.7 (q)       |                        |
| 6″           |                | 20.7 (q)       |                        |
|              |                |                |                        |

Table 1. <sup>13</sup>C NMR data of antiostatins  $A_1$  and  $B_4$ , and carazostatin<sup>a</sup>.

- <sup>a</sup> Solvent:  $A_1$  and  $B_4$  in  $(CD_3)_2CO$ , carazostatin in  $CDCl_3$ .
- <sup>b</sup>  $q = CH_3, t = CH_2, d = CH, s = -C.$

with a carbazomycin related compound carazostatin<sup>7</sup>) (Fig. 2).

Detailed analysis of the <sup>1</sup>H NMR spectral data (500 MHz, acetone- $d_6$ ) of antiostatin A<sub>1</sub> revealed the presence of an n-pentyl side chain (CH<sub>3</sub>, t;  $0.89 \text{ ppm}; 3 \times \text{CH}_2$ , each m, 1.37, 1.45 and 1.65 ppm; CH<sub>2</sub>, t, 2.97 ppm), an N-acetyl group (CH<sub>3</sub>, s, 2.47 ppm, NH, br s, 9.68 ppm), an aromatic methyl group (CH<sub>3</sub>, s, 2.40 ppm), a 1,2-disubstituted benzene ring (8.13, 7.10, 7.30 and 7.45 ppm), a phenolic hydroxy group (s, 8.05 ppm) and an imino proton of a carbazole nucleus (brs, 10.16 ppm). Based on heteronuclear multiple-bond correlation (HMBC)8) spectral analysis (vide infra) and comparison of the <sup>13</sup>C NMR spectral data (125 MHz, acetone-d<sub>6</sub>) of antiostatin  $A_1$  and carazostatin (Table 1), these groups were arranged on the carbazole nucleus as shown in Fig. 1.

Thus, the methylene proton (1'-H, 2.97 ppm) of the *n*-pentyl side chain was coupled to C-9a (135.1 ppm), C-1 (123.3 ppm) and C-2 (125.8 ppm). The aromatic methyl proton was coupled to the last Fig. 1. Partial structure of antiostatin  $A_1$  as revealed by HMBC analysis.

Arrows indicate <sup>13</sup>C-<sup>1</sup>H long range couplings.



- Fig. 2. Structures of antiostatins A series and carazostatin.
  - (a) Antiostatin series:  $A_1$ ,  $R = (CH_2)_4CH_3$ ;  $A_2$ ,  $R = (CH_2)_2CH(CH_3)CH_2CH_3$ ;  $A_3$ ,  $R = (CH_2)_4CH(CH_3)_2$ ;  $A_4$ ,  $R = (CH_2)_6CH_3$ . (b) Carazostatin,  $R = (CH_2)_6CH_3$ .



two carbons and C-3 (144.3 ppm), which in turn was coupled to the hydroxy proton and amide proton. Additional <sup>13</sup>C-<sup>1</sup>H couplings observed between the amide carbonyl carbon (C-1") and acetyl methyl and amide protons together with the data just described corroborated the relationship among C-1 to C-4 of the carbazole nucleus as well as the substituents on these carbons as shown in Fig. 1. Thus, the structure of antiostatin A<sub>1</sub> has been determined to be 1-*n*-pentyl-2-methyl-3-hydroxy-4-acetylaminocarbazole (Fig. 2).

The <sup>1</sup>H and <sup>13</sup>C NMR spectral comparison of antiostatin  $B_4$  and antiostatin  $A_1$  showed that the acetyl group in  $A_1$  was substituted by an isobutylamino group (-NH-CH<sub>2</sub>, 3.19 ppm, CH, 1.88 ppm, 2 × CH<sub>3</sub>, 0.98 ppm) in  $B_4$  in addition to the replacement of the alkyl side chain in  $A_1$  by an *n*-heptyl residue (CH<sub>3</sub>, t, 0.89 ppm, 5 × CH<sub>2</sub>, m, 1.28, 1.30, 1.36, 1.47 and 1.66 ppm, CH<sub>2</sub>, t, 2.98 ppm) in  $B_4$  (Fig. 3).

In the HMBC spectrum of antiostatin  $B_4$ , the methylene proton (1'-H, 2.98 ppm) of the *n*-heptyl chain was coupled to C-1 (123.7 ppm), C-2 (126.0 ppm) and C-9a (135.3 ppm). The aminomethylene proton of the isobutylamino group (3"-H, 3.19 ppm) was coupled to a carbonyl carbon

Fig. 3. Partial structure of antiostatin  $B_4$  as revealed by HMBC analysis.

Arrows indicate <sup>13</sup>C-<sup>1</sup>H long range couplings.





 $\begin{array}{ll} B_2, & R = (CH_2)_5 CH_3; & B_3, & R = (CH_2)_4 CH(CH_3)_2; \\ B_4, & R = (CH_2)_6 CH_3; & B_5, & R = (CH_2)_5 CH(CH_3)_2. \end{array}$ 



(C-2", 156.8 ppm) indicating the presence of a unit  $(CH_3)_2 CH - CH_2 - NH - C = O$ . Thus, there remained one carbonyl or amidino carbon (155.9 ppm), two exchangeable protons (6.92 and 10.90 ppm), one oxygen and two nitrogens to be explained. Since the carbon chemical shifts of C-4 in  $B_4$  and  $A_1$  were almost identical (117.6 vs. 118.3 ppm), C-4 was substituted by a nitrogen atom. Therefore, the functional group at C-4 in  $B_4$  was either  $C_4 - NH - C(=O) - NH - C(=O) - O(=O) - O(=$  $NH - C_4H_9$  or  $C_4 - NH - C(=NH) - O - C(=O) - O_{-}C(=O)$  $NH-C_4H_9$ . This final problem was solved by the fragmentation ions observed in the HRFAB-MS at 336.1846 (C21H24N2O2, calcd 336.1838) and 251.0826 ( $C_{21}H_{24}N_2O_2 - C_6H_{13}$ , calcd 251.0821) as shown in Fig. 3. Based on these results, the structure of antiostatin  $B_4$  has been determined as shown in Fig. 4.

Because of the paucity of the samples available, the structures of the remaining components, antiostatins A<sub>2</sub>, A<sub>3</sub>, A<sub>4</sub> (Fig. 2), B<sub>2</sub>, B<sub>3</sub> and B<sub>5</sub> (Fig. 4) were established by comparison of <sup>1</sup>H NMR and MS spectral data. Their molecular formulae and mp's are as follows; A<sub>2</sub> (yield 0.8 mg) C<sub>21</sub>H<sub>26</sub>N<sub>2</sub>O<sub>2</sub>, 195~197°C; A<sub>3</sub> (0.75 mg) C<sub>22</sub>H<sub>28</sub>N<sub>2</sub>O<sub>2</sub>, 190~ 192°C; A<sub>4</sub> (1.5 mg) C<sub>22</sub>H<sub>28</sub>N<sub>2</sub>O<sub>2</sub>, 191~193°C; B<sub>2</sub> (0.5 mg) C<sub>25</sub>H<sub>34</sub>N<sub>4</sub>O<sub>3</sub>, 119~120°C; B<sub>3</sub> (1.3 mg)  $C_{26}H_{36}N_4O_3$ ,  $117 \sim 118^{\circ}C$ ;  $B_5$  (1.2 mg)  $C_{27}H_{38}N_4O_3$ ,  $92 \sim 94^{\circ}C$ .

Antiostatins are the first carbazole derivatives that possess an acetamide group or a substituted urea chain in addition to a long alkyl chain. The IC<sub>50</sub> values of antiostatin A<sub>1</sub> and antiostatin B<sub>4</sub> for the assay system mentioned above were 0.207 and 0.211  $\mu$ g/ml, respectively, while that of vitamin E was 10.8  $\mu$ g/ml. The remaining members of the antiostatins showed similar IC<sub>50</sub> values to those of A<sub>1</sub> and B<sub>4</sub>.

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